

ATTORNEY DOCKET NUMBER: 7853-240  
SERIAL NUMBER: TBA  
REFERENCE: AD

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BLOOD PLATELET LYSATE, METHOD OF ITS PREPARATION  
AND A CELL CULTURE MEDIUM CONTAINING SAID BLOOD PLATELET  
LYSATE

The present invention relates to a blood platelet lysate obtained from plasma from animal whole blood, a method for preparing the blood platelet lysate and a cell culture medium containing the blood platelet lysate.

5       A problem in the slaughterhouse industry in an international perspective is, in alia, to find a continuous and profitable market for slaughter blood. In many countries, the blood is even discarded as waste material for lack of possibilities of processing or  
10       using the blood. In cases where the slaughter blood is taken care of, it is mainly used as an admixture to foodstuffs and animal feed. The hemoglobin part of the blood can be used for different blood products, such as black-pudding, and the plasma part can be used as a  
15       thickener, a protein-enriching agent etc. Yet there are not always marketing possibilities for all slaughter blood, and it is therefore important to find new fields of application.

20       In biotechnology, a further problem is that along with the ever increasing culture of mammalian cells and other animal cells for production and research, one of the main components of the growth medium, foetal calf serum (FCS), has become a limiting factor. Foetal calf serum (FCS) is extracted from calf foetus from  
25       slaughtered cows. There are also other drawbacks: FCS is expensive, and the quality when culturing a special cell line may vary strongly from batch to batch. Large efforts have therefore been made to develop alternatives to FCS.

30       We have now surprisingly found that the blood platelet fraction in the plasma part in blood is an excellent agent for cell culture, and that it can wholly

or partly replace FCS, especially when culturing hybridoma cells.

Many different attempts to find alternatives to FCS are known. In US patent specification 4,350,687 the  
5 object is to produce a single, simple and well-defined component to promote the growth of tumor cells. The source is platelets from human blood which is outdated and has become unfit for clinical use. This patent  
10 specification also states that extracts from platelets can promote the growth of 3T3 cells, SV3T3 cells and mouse fibroblasts which have been transformed. Gospadoro-witz and Moran (Ann. Rev. Biochem. 45 (1976) 540) pointed out that the mitogenic factor for 3T3 cells should be present in platelets.

15 Platelet extracts are also known to stimulate primary cultures from pigs ("Serum-free medium enhances growth and differentiation of cultured pig granulosa cells" Barano J L S; Hammond J M, Milton S, Endocrinology 116 (1), 1985, 51-58), and "skin explants" ("Support  
20 and stimulation of epidermal cell outgrowth from porcines skin explants by platelet factors". Hebda PA, Alstadt SP, Hileman WT, Eaglstein WH. Br J Dermatol 1986 Nov; 115(5):529-41).

According to the classical theory, the role of the  
25 platelets is to aggregate on the site of the injury, to form a plug and to empty their contents of growth factors and other mediators, so as to stimulate connective tissue to grow, migrate and heal the wound, and to stimulate vascularisation.

30 A further attempt is stated in European patent specification 0,104,831. Here the starting material is material that normally goes to waste in slaughter, viz. "tissue juices together with flowing blood, a semi-solid, rubbery mass". The efficiency of this product is again  
35 demonstrated on cells which can be expected to respond in accordance with the classical theory: WI-38 (human fibroblasts); and on cell line VerO from African monkey

kidney (transformed cells).

These previous attempts have all concerned either transformed cells or cells on which platelets can act in accordance with the classical theory.

5       The idea that platelets can play a role other than indicated by the classical theory, was recently suggested (Page, C.P. TIPS 9 (1988) 66-71). However, nowhere in this paper is it indicated that platelets or their contents stimulate the growth and division of cells  
10       which have been obtained from the immunological system.

      The present invention is based on the unexpected and surprising result that a platelet extract stimulates the growth and division of cells from the immunological system, viz. hybridoma cells in which the original two  
15       components - the B-cell and the leukaemia cell - are both derived from the immunological system. Moreover, it is shown that the production of monoclonal antibodies in hybridoma cells which have been cultured in the product according to the present invention, may be well compared  
20       with the one obtained when hybridoma cells are cultured in FCS. Finally, the present invention shows that the platelet product functions poorly for a number of transformed cells, which goes to show that the demonstrated effect cannot be a general growth-promoting  
25       effect of platelets on transformed cells.

      Common substitutes for FCS are serum-free, synthetic media containing various growth factors and, in many cases, e.g. transferrin and insulin. When FCS is used, the added amount is usually between 1 and 20%, in many  
30       cases 10% (% by volume). The remaining component is a nutrient medium containing salts and amino acids. On an industrial scale, serum-free media are frequently preferred for economical reasons.

      One object of the present invention is to provide a  
35       concentrated platelet lysate which has been obtained from plasma from animal whole blood, young as well as adult animals, and which wholly or partly can replace FCS

in cell culture media.

A further object of the present invention is to provide a method for preparing said platelet lysate.

5 A still further object of the present invention is to provide a cell culture medium containing said platelet lysate, a conventional nutrient medium and, optionally, FCS.

10 According to the present invention, a platelet lysate for use in cell culture is provided, characterised in that it is prepared from plasma from whole blood from slaughter animals, and that it consists mainly of lysed platelets.

15 Furthermore, there is provided a method for preparing a platelet lysate derived from plasma from animal whole blood to which a citrate has been added to avoid coagulation, the plasma being of food quality and having a hemolysis figure of about 1-3, characterised in

- a) that said plasma is centrifuged so as to obtain a platelet-rich paste,
- 20 b) that  $\text{Ca}^{2+}$  is added to said paste, in equimolar amounts in relation to the citrate content in the original plasma, for lysing the included platelets and coagulating the remaining components, a clear, light red liquid of lysate which to a substantial degree consists of lysed platelets, and a
- 25 c) that said clear, light red liquid is sterile-filtered, and
- d) that the platelet lysate thus obtained is collected.

30 Further there is provided a cell culture medium characterised in that it contains

- a) a platelet lysate which has been prepared from plasma from whole blood from slaughter animals and consists mainly of lysed platelets,
  - 35 b) a conventional nutrient substrate, and
  - c) optionally, foetal calf serum,
- and use thereof for culture of hybridoma cells.

The animals which are intended according to the present invention can in principle be all animals, young or adult, from which the required amount of platelets can reasonably be collected. Use is preferably made of blood from mainly adult animals, since this is more economical. Examples of suitable animals are slaughter animals and other farmed animals such as cattle, pigs, sheep or poultry. Preferably, use is made of whole blood from cattle or pigs, most advantageously cattle.

The cell culture medium according to the present invention has appeared to be especially useful for culturing hybridoma cells which are today used to produce monoclonal antibodies. The present cell culture medium can, wholly or partly, replace foetal calf serum.

Below follows a detailed description of preferred embodiments of the present invention.

For preparing the present platelet lysate, use is made of whole blood from animals, such as slaughter cattle or pigs. The animals should be healthy and satisfy the requirements that are put forward in veterinary inspection of animals intended for foodstuffs according to Swedish regulations, as well as under EEC and US rules.

After drawing off, the blood is cooled rapidly, preferably to about +4°C within a minute. A citrate, such as sodium citrate, is added to prevent coagulation. Optionally, the blood is then stored in cooling tanks until it is time for further treatment.

Subsequently, the blood is examined, both in bacteriological and in sensory respect. The blood should preferably satisfy the requirements which the National Food Administration places on products intended for foodstuffs, and moreover the blood should satisfy the corresponding regulations within the EEC and the US. From experience we have found that suitable upper limits for the bacterial content in blood are those stated in

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Table I below. The sensory analysis is carried out as follows: one smells the blood and by experience decides whether the blood is acceptable or not.

Bacteriological samples are preferably taken  
5 both from whole blood and from separated plasma.

TABLE I

10	BACTERIOLOGICAL LIMIT OF ACCEPTANCE	
	<u>Number of bacteria/ml</u>	
	Aerobic number of bacteria (tryptonglucose extract agar 30° for 3 days)	< 100,000
15	Thermostable coli (VGR-agar 44° for 1 day)	< 300
20	Tellurite-resistant enterococci (Slanetz-Bartley agar 37° for 2 days)	< 3,000
	DNAs-positive staphylococci (Baird-Parker agar 37° for 2 days)	< 300
25	Occurrence of salmonellae (Enrichment broth-SS agar)	negative

Based on the results from examination and analysis,  
30 it is decided whether the blood can be used or not. The  
blood is separated into plasma and red blood corpuscles.  
Use is preferably made of a blood separator, but also  
other suitable, traditional separators can be used.  
All the time the blood is kept cool, preferably at about  
35 +4°C. From the whole blood, a separate sample is taken  
for hemolysis assessment of the plasma part. The hemo-  
lysis figure is a measure of the amount of lysed red



blood corpuscles. A preferred method of measuring the hemolysis figure is ocular assessment of the colour of the plasma by comparison with a colour chart. The colour values on this chart are shown in Table II.

- 5 Colour measuring is suitably made on samples after table centrifuging in test tubes. The plasma used in the present method has a hemolysis figure of 1-3, preferably 1-2 and most preferred 1.

10

TABLE II

COLOUR MEASURING OF BOVINE PLASMA (PHOTO)			
15 Hemolysis figure	L* (lightness)	a* (redness)	b* (yellowness)
1	35.44	13.20	39.12
2	26.98	18.85	26.14
3	21.59	20.27	16.58
20 4	17.91	22.09	11.54
5	16.32	19.83	7.35
6	12.74	15.47	4.09
7	11.52	9.69	1.67
8	11.59	6.50	0.23
25			

L\* = lightness; lighter the higher the value

a\* = redness; redder the higher the value

b\* = yellowness; more yellow the higher the value

30

Instruments and measuring conditions:

Hunterlab Color Quest

Colour chart: CIELAB (1976)

Illuminant D65, 10° standard observer, specular excluded

Measuring opening: 6 mm diameter, white background

35

Then the platelet fraction was concentrated from the plasma. This is preferably carried out in that the plasma is first centrifuged to about 100-fold con-

centration, and subsequently this concentrate is further concentrated by a second centrifugation. Now a solid phase of concentrated platelets is obtained. To this solid phase there are added  $\text{Ca}^{2+}$  ions, for example  
5 in the form of  $\text{CaCl}_2$ , in an equimolar amount in relation to the citrate content of the original plasma. The addition of  $\text{Ca}^{2+}$  causes fibrinogen etc to coagulate and the platelets to lyse, which gives a clear, light red top liquid. This clear liquid is sterile-filtered, and  
10 the platelet lysate is ready for use.

The method can be a continuous method, a batchwise method or a combination of these. If desired, several concentration steps, such as a centrifugation step, a filtration step or some other analogous step, can  
15 be introduced.

The platelet lysate according to the present invention consists mainly of lysed platelets, especially about 90-100% by volume, most preferred about 95-100% by volume.

20 The cell culture medium according to the present invention contains, in addition to the present platelet lysate, a nutrient medium containing salts etc and, optionally, FCS. The nutrient medium can be any suitable, conventional nutrient medium, e.g. one stated in the  
25 Sigma catalogue issued by Sigma Chemical Company.

The amount of FCS which is optionally added is not critical, but is selected as required and desired. For economical reasons, it is frequently desired to minimise the amount of FCS used, but for different other  
30 reasons, an addition may be required.

For the purpose of further describing the advantages and purposes of the present invention, a number of non-limiting Examples are stated below.

In all Examples, the platelet lysate according  
35 to the present invention is designated "S". The FCS used has been supplied by Gibco.

Example 1 states a preferred method for preparing

a platelet lysate according to the present invention.

EXAMPLE 1

From a continuous process for separating plasma from whole blood from slaughter cattle, about 1 ton of plasma was taken. A bacteriological examination of the whole blood yielded the following values:

A sensory and bacteriological analysis yielded an acceptable result. The yield of plasma from whole blood is about 60%. Thus, the amount of plasma obtained corresponds to about 1.7 ton of blood. The plasma had a hemolysis figure of 2. A bacteriological examination was also made of the plasma which was found to be acceptable.

First the plasma was centrifuged in a so-called bactofuge (Alfa Laval type D 3187 M) for 15 minutes. 5734 g of cell-rich plasma was obtained, which was centrifuged at 925 g in a Mistral 6 l at 4°C. The obtained gel-like paste, 127 g, which was white to red coloured, was collected.

63.5 g of water was added to the paste (1:0.5, weight:weight, paste:water) and the mixture was stored in a refrigerator at 4°C for 15 minutes. After 15 minutes, 127 g of 0.024 M  $\text{CaCl}_2$  was added (equimolarly in relation to the citrate content in the blood), and the sample was centrifuged directly at 1900 g in a Mistral 6 l at 4°C.

The supernatant which was red to whitish, was collected and combined with the liquid that was included in the paste after the second centrifugation at 1900 g. The liquid from the paste was extracted in that the paste was transferred to a Büchner funnel with a fixed sieve plate, and was agitated by a spoon until all liquid had been extracted. The total amount of liquid was 176 g.

The total volume of liquid (176 g) was then filtered through two Millipore filters; first through a filter under the name of Milligord TP, catalogue No. CW SS 004 T4, and then through a sterile filter, Millipack <sup>TM</sup> 200, catalogue No. MPGL 20C A3. Before filtration, the filters

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were soaked with distilled water. Total amount of filtrate: 157 g.

The filtration was carried out between 0.5 and 4 bar. The sterile liquid which is the final product, was  
 5 filtered down into sterile polypropylene flasks Nalge 2106, and the flasks were stored at 4°C.

Table III presents a sampling schedule for the experiment, and Table IV gives the results of analysis in the analysis positions as stated. The analyses have  
 10 been carried out at the Agricultural University of Sweden, Faculty of Veterinary Medicine, Institute of Clinical Chemistry, P.O. Box 7038, 750 07 Uppsala.

TABLE III

15

SAMPLING SCHEDULE FOR THE ANALYSIS		
		<u>Analysis position</u>
20	Whole blood	1
	Centrifugation	
	Plasma	2
	Bactofugation	
	Cell-rich plasma	3
25	Paste	Centrifugation 925 g
	Lysing H <sub>2</sub> O CaCl <sub>2</sub>	
	-	4
	Centrifugation 1900 g	
30	Supernatant	5
	Filtration	
	Sterile product	6

35

TABLE IV

5	RESULT OF THE ANALYSIS				
	Analysis position	Blood leucocytes Conc. of particles $\times 10^9/L$	Platelets Conc. of particles $\times 10^9/L$	Erythrocytes Conc. of particles $\times 10^{12}/L$	Protein % by weight
10					
	1	5.6	296	7.85	17.9
	2	0.3	154	0.02	7.1
	3	0	370	0	6.2
15	4	0	24	0	3.8
	5	0	15	0	2.0
	6	0	12	0	Not analysed

20        The method can be optimised by e.g. further centrifugation and filtration so as to increase the yield and, quality of the final product. Also other steps of concentrating which can be compared to centrifugation and filtration and are based on, for example, the affinity principle, may be used.

25        Examples 2 and 3 below concern culture experiments with hybridoma cells, in which the adding of FCS only, (for comparison purposes) to a nutrient medium (Dulbecco's min. medium, Gibco) has been compared with the adding of  
30        a mixture of FCS and the present platelet lysate in different percentages.

      The concentrations of FCS or FCS+S are stated in % by volume of the total volume of cell culture medium.

#### EXAMPLE 2

35        8F4 hybridoma cells were cultured ( $37^\circ C$  in 8%  $CO_2$ ), and the cells were counted in a Bürker chamber on

12

day 2, 3, 4, 5 and 6.

The cells have first been adapted to the S-supplement by being cultured for about 2 weeks in a medium containing 1% FCS + 1% S-supplement. The experiments were here carried out in single samples.

		Number of cells per ml x 10 <sup>6</sup>					
		FCS 10%	FCS 1%	FCS 5%	FCS+S 1+5%	FCS+S 1+1%	FCS+S 1+0.2%
10							
	8F4 hybridoma						
	Day 2	0.72	0.36	0.38	0.59	0.46	0.40
	Day 3	1.38	0.61	0.64	1.06	0.78	0.62
	Day 4	1.93	0.94	1.12	1.52	1.04	0.92
15	Day 5	2.88	1.11	1.28	1.72	1.33	1.08
	Day 6	2.51	1.22	1.58	1.98	1.22	1.26

### EXAMPLE 3

20 SK4H hybridoma cells were cultured and counted as in Example 2.

		Number of cells per ml x 10 <sup>6</sup>					
25		FCS 10+%	FCS 1%	FCS 5%	FCS+S 1+5%	FCS+S 1+1%	FCS+S 1+0.2%
SK4H hybridoma							
	Day 2	0.86	0.30	0.52	0.61	0.43	0.27
	Day 3	2.70	0.82	1.50	1.83	1.24	1.06
30	Day 4	3.58	1.06	1.90	2.72	2.00	1.48
	Day 5	3.34	1.44	2.05	2.52	1.97	1.70
	Day 6	2.24	1.26	1.54	2.11	1.03	1.18

35 Examples 2 and 3 clearly show that the platelet lysate according to the present invention is an excellent substitute for FCS. As appears, an admixture of 10% FCS

can substantially be replaced by an admixture of 1% FCS and 5% of the present platelet lysate.

Examples 4, 5 and 6 concern culture experiments with 8F4 hybridoma cells in Dulbecco's min. medium (Gibco) with additions of FCS, the present platelet lysate (S) or mixtures thereof. The present platelet lysate has been inactivated at 56°C for 30 minutes and, respectively, has not been inactivated. The cells have been placed in cell culture plates, in an original concentration of 50,000 cells/ml (Examples 4 and 5) or in culture flasks in a cell concentration of 75,000 cells /ml (Example 6).

#### EXAMPLE 4

	Inactivated S					
	FCS 10%	FCS 1%	S 15%	S 5%	FCS+S 1+5%	FCS+S 1+1%
Day 2	0.232	0.072	0.177	0.120	0.290	0.960
Day 3	0.590	0.085	0.430	0.195	0.464	0.180
Day 4	1.205	0.155	0.665	0.375	0.915	0.285
Day 5	1.510	0.250	1.525	0.515	1.045	0.480
Day 6	1.355	0.315	1.590	0.695	0.965	0.280

#### EXAMPLE 5

	Not inactivated S					
	FCS 10%	FCS 1%	S 15%	S 5%	FCS+S 1+5%	FCS+S 1+1%
Day 1	0.056	0.037	0.060	0.036	0.072	0.064
Day 2	0.236	0.076	0.280	0.130	0.230	0.112
Day 3	0.680	0.100	0.450	0.280	0.485	0.200
Day 4	0.985	0.155	0.630	0.370	0.815	0.280
Day 5	1.695	0.200	1.290	0.675	1.020	0.590
Day 6	1.405	0.280	1.390	0.775	0.965	0.445

EXAMPLE 6

Culture in flasks								
	FCS 10%	FCS 1%	FCS+S 1+15*	FCS+S 1+5*	FCS+S 1+1*	FCS+S 1+15	FCS+S 1+5	FCS+S 1+1
Day 4	1.76	0.19	1.47	1.07	0.48	1.68	1.06	0.55
Day 5								
Day 6	2.52	0.32	2.17	1.68	0.73	2.01	1.64	0.76

\* ) inactivated

As appears again from Examples 4-6, 1% FCS + 5% S constitute an acceptable alternative to 10% FCS. Moreover, 15% S can completely replace 10% FCS. It further appears that an inactivation of S does not affect the growth of cells.

EXAMPLE 7

Culture experiments with FCS and S (inactivated), respectively, on 2F6 hybridoma cells. The cells were placed in an original concentration of 50,000 cells/ml.

	FCS 10%	FCS 1%	S 15%	S 5%	FCS+S 1+5%	FCS+S 1+1%
Day 1	0.049	0.027	0.042	0.044	0.052	0.040
Day 2	0.173	0.041	0.136	0.109	0.157	0.092
Day 3	0.433	0.089	0.339	0.232	0.301	0.156
Day 4	0.890	0.092	0.707	0.409	0.583	0.238
Day 5	1.473	0.111	1.386	0.602	0.975	0.440
Day 6	1.660	0.158	1.775	0.810	1.132	0.414

This Example again shows that 1% FCS + 5% S can substantially replace 10% FCS. This also applies to 15% S.

Examples 8, 9 and 10 concern the production of anti-bodies (Ab) from hybridoma cells in different concentra-



15

tions of FCS, S and, respectively, mixtures thereof.

Experimental conditions: ELISA plates were coated with sorbent-purified anti-mouse IgG antibodies (10 µg/ml), and hybridoma supernatants were added in serial dilution.

5 Bound mouse antibodies were detected by addition of ALP-conjugated anti-mouse IgG and substrates (paranitro-phenyl sulphate). The experiments were carried out in duplicate samples, and determinations of the concentration were obtained by comparison with a known standard  
10 of mouse IgG cord parallel in each test run.

#### EXAMPLE 8

Production of antibodies from 8F4 hybridoma cells cultured in different concentrations of FCS and S. The supernatants on day 5 were measured.

15

Test	Hyb/Day	FCS	S	Amount of Ab (µg/ml)
1	8F4 5	10%	-	10.62
20 2	8F4 5	1%	-	3.25
3	8F4 5	-	5%	10.00
4	8F4 5	1%	5%	11.50
5	8F4 5	1%	1%	5.47
6	8F4 5	1%	0.2%	4.53

25

#### EXAMPLE 9

Production of antibodies from 8F4 hybridoma cells cultured in different concentrations of FCS and S.

30 Supernatants on day 6 were measured.

	Test	Hyb/Day	FCS	S	Amount of Ab ( $\mu\text{g/ml}$ )
5	1	8F4 6	10%	-	6.25
	2	8F4 6	1%	-	1.56
	3	8F4 6	1%	15% ia	7.44
	4	8F4 6	1%	5% ia	8.12
	5	8F4 6	1%	1% ia	2.44
10	6	8F4 6	1%	15%	8.15
	7	8F4 6	1%	5%	8.44
	8	8F4 6	1%	1%	2.06

ia = inactivated at 56°C for 30 minutes.

15 The tests show a slightly higher production of antibodies in cultures with 1% FCS + 5% S as compared to cultures with 10% FCS.

#### EXAMPLE 10

20 Production of antibodies from SK4H hybridoma cells in FCS and S. The supernatants on day 5 were measured, but supernatants from day 3 indicate a similar tendency, but with lower total concentrations.

	Test	Hyb/Day	FCS	S	Amount of Ab ( $\mu\text{g/ml}$ )
25	1	SK4H 5	10%	-	13.75
	2	SK4H 5	1%	-	9.63
	3	SK4H 5	-	5%	17.31
	4	SK4H 5	1%	5%	20.78
30	5	SK4H 5	1%	1%	Not assessable
	6	SK4H 5	1%	0.2%	12.50

35 The SK4H hybridoma produced slightly higher amounts of antibody, which conforms to previous observations made on several occasions. Here, too, the platelet lysate (S) according to the present invention (5%)

seems to have a favourable effect on the production of antibodies as compared to culture in FCS only.

Since the present platelet lysate can be obtained from slaughter animals, both adult and young animals, essential advantages in respect of economy and availability are obtained.

Thus, it has surprisingly appeared that the hybridoma cells grow and produce antibodies in a most satisfactory fashion in the present platelet lysate.

Different types of cell necessitate different specific conditions of culture, which means that it is not possible to safely determine whether a certain type of cell grows in a satisfactory manner in a certain type of medium.

In order to elucidate this, experimental cultures of different tumour cells were made in a nutrient medium to which FCS and the present platelet lysate (S) were added. These experiments are accounted for in Examples 11 and 12.

#### EXAMPLE 11

HL-60 cells (Human promyelocytic leukaemia) were cultured in a suspension culture with the nutrient medium RPMI 1640 (Gibco) + antibiotics (penicillin 50 IU/ml and streptomycin 50 µg/ml) and 10% FCS. The cultures were incubated at 37°C in humid atmosphere of 5% CO<sub>2</sub> in air. When the cells had reached the flat phase, a new culture (1st passage) was made with 10% of the platelet lysate (S) according to the present invention under the same conditions, and then a 2nd passage with 10% S. The cells were counted in a Bürker chamber. The results are stated in Fig. 1 in which an unfilled ring (○) concerns experiments with 10% FCS, a filled ring (●) the 1st passage with 10% S, and a filled square (■) the 2nd passage with 10% S. All points constitute an average value of two measurements.

The results show quite clearly that the platelet

lysate according to the present invention is not suited as a medium in culturing this type of tumour cells.

EXAMPLE 12

5 In this experiment, L1210<sub>D20</sub> (DL- $\alpha$ -difluoromethyl-ornithine-resistant mouse-leukaemia cell line) was cultured in a suspension culture with the same nutrient medium as in Example 11 plus 50  $\mu$ M  $\beta$ -mercaptoethanol and 20 mM  $\alpha$ -difluoromethylornithine.

10 The results are stated in Fig. 2 in which the same designations are used as in Example 11. Only the 1st passage with 10% S was carried out. Nor did this cell line grow acceptably in the presence of the platelet lysate according to the present invention.

15 According to the present invention, there is thus provided a platelet lysate for cell culture, said platelet lysate being able to replace, wholly or partly, foetal calf serum and, accordingly, considerably improve the economy and availability of the culture medium.

20 According to the present invention, also a new and profitable use of animal whole blood is obtained.

## CLAIMS

1. A blood platelet lysate for use in cell culture, characterised in that it has been prepared from plasma from animal whole blood, and that it consists mainly of lysed platelets.

5        2. The blood platelet lysate according to claim 1, wherein it consists of lysed platelets to about 90-100% by volume, most preferred 95-100% by volume.

3. The blood platelet lysate according to claim 1 or 2, characterised in that said plasma is of food quality, and that it has a hemolysis figure of about 1-3, preferably 1-2.

10       4. A method for preparing a blood platelet lysate derived from plasma from animal whole blood to which a citrate has been added to avoid coagulation, characterised in

- 15       a) that said plasma is centrifuged so as to obtain a platelet-rich paste,
- 20       b) that  $\text{Ca}^{2+}$  is added to said paste, in equimolar amounts in relation to the citrate content in the original plasma, for lysing the included platelets and coagulating the remaining components, a clear, light red liquid of lysate which to a substantial degree consists of lysed platelets, and a coagulate being obtained,
- 25       c) that said clear, light red liquid is sterile-filtered, and
- d) that the platelet lysate thus obtained is collected.

5. The method according to claim 4, wherein the platelet lysate consists of lysed platelets to about 90-100% by volume, most preferred 95-100% by volume.

6. The method according to claim 4 or 5, wherein said plasma is of food quality and has a hemolysis figure of 1-3, preferably 1-2.

7. The method according to any one of claims 4-6, wherein in step a), said plasma is centrifuged in a bactofuge to yield about 100-fold concentration, and that this concentrate is then centrifuged a second time  
5 to yield a further concentration.

8. A cell culture medium c h a r a c t e r i s e d in that it contains

- a) a platelet lysate which has been prepared from plasma from animal whole blood and con-  
10 sists mainly of lysed platelets,
- b) a conventional nutrient substrate, and
- c) optionally, foetal calf serum.

9. A cell culture medium c h a r a c t e r i s e d in that it contains

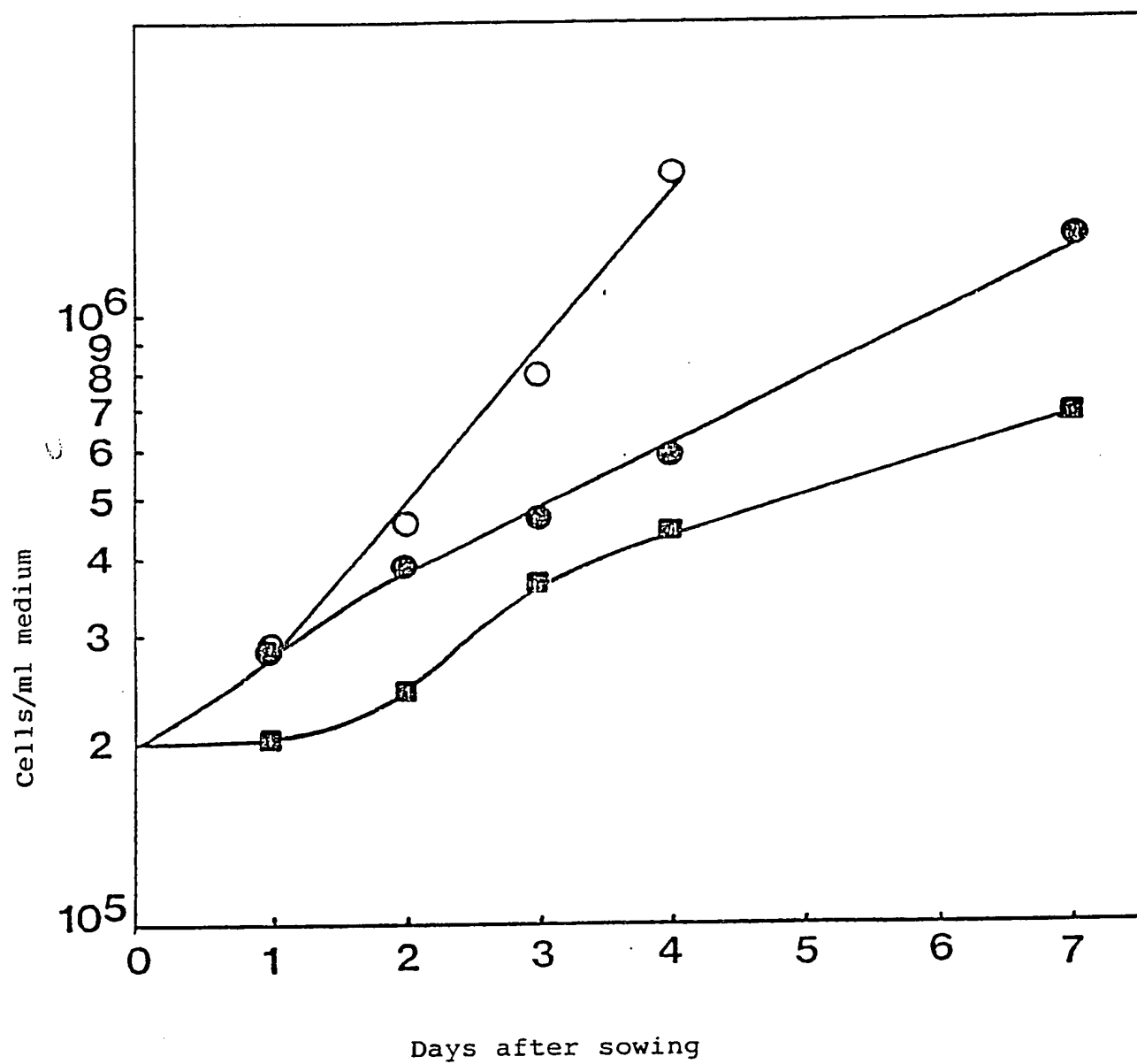
- 15 a) a platelet lysate which has been prepared by the method according to claims 4-7,
- b) a conventional nutrient substrate, and
- c) optionally, foetal calf serum.

10. Use of the cell culture medium according  
20 to claim 8 or 9 for culture of hybridoma cells.

11. A method for culture of hybridoma cells, c h a r a c t e r i s e d in that said culture is carried out in a medium according to claim 8 or 9.

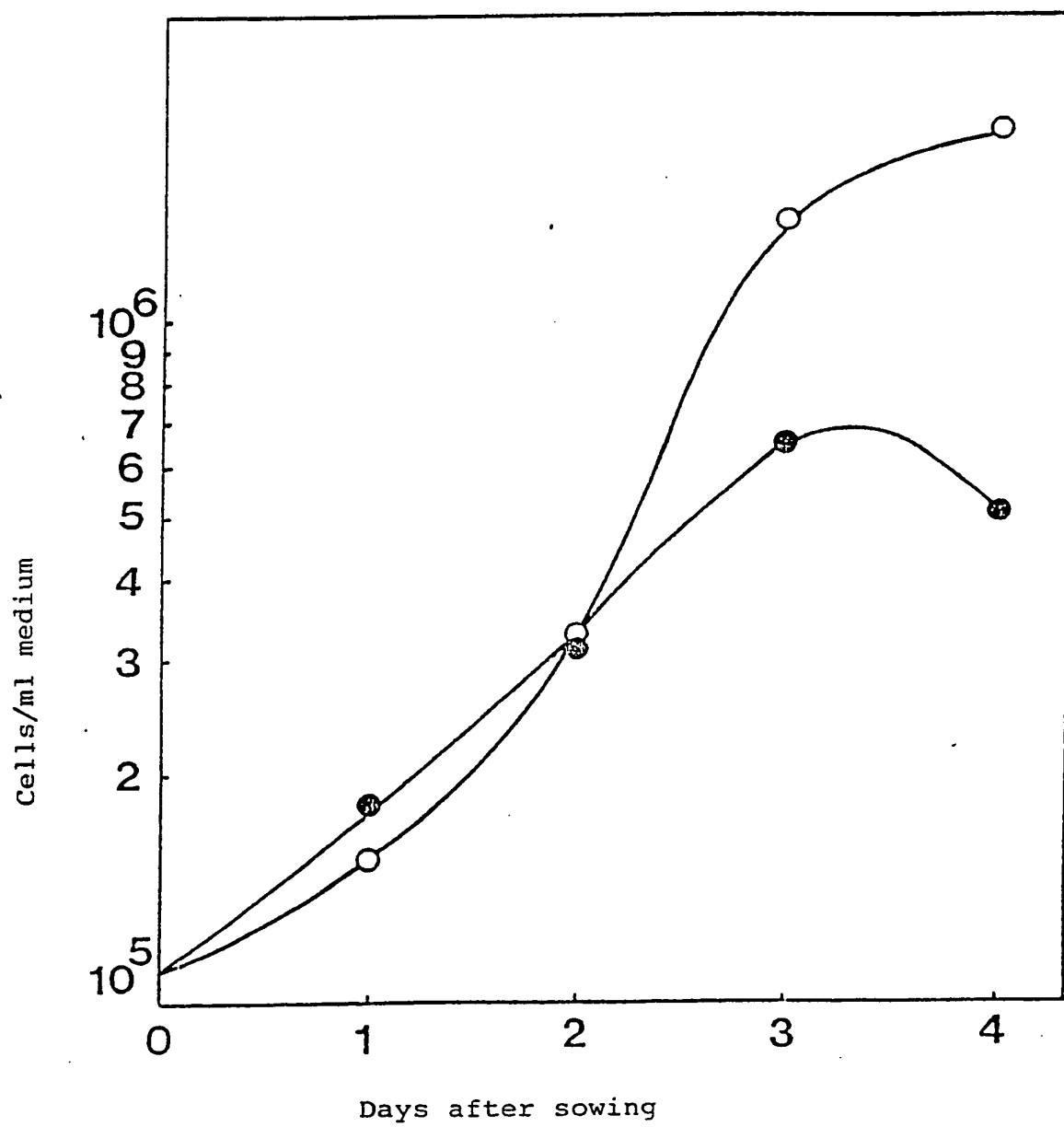
1/2

FIG. 1



2/2

FIG. 2





# INTERNATIONAL SEARCH REPORT

International Application No PCT/SE89/00232

<b>I. CLASSIFICATION F SUBJECT MATTER</b> (if several classification symbols apply, indicate all) <sup>9</sup> According to International Patent Classification (IPC) or to both National Classification and IPC 4 <div style="margin-top: 10px;">C 12 N 5/00</div>																						
<b>II. FIELDS SEARCHED</b> <div style="text-align: center; margin-top: 10px;">Minimum Documentation Searched <sup>7</sup></div> <table border="1" style="width: 100%; border-collapse: collapse; margin-top: 5px;"> <tr> <th style="width: 50%; padding: 5px;">Classification System</th> <th style="width: 50%; padding: 5px;">Classification Symbols</th> </tr> <tr> <td style="padding: 5px;">IPC 4</td> <td style="padding: 5px;">C 12 N</td> </tr> <tr> <td style="padding: 5px;">US C1</td> <td style="padding: 5px;">435</td> </tr> </table> <div style="text-align: center; margin-top: 10px;">Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched <sup>8</sup></div> <div style="margin-top: 20px;">SE, NO, DK, FI classes as above</div>			Classification System	Classification Symbols	IPC 4	C 12 N	US C1	435														
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## III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No
X	Chemical Abstracts, Vol. 95, No. 19, 9 November 1981, (Columbus, Ohio, US), see page 122, abstract 162602m, P.D. Phillips et al.: "Growth regulation of WI138 cells in a serum-free medium", & Exp. Cell Res. 1981, 134(2), 297-302 (Eng).	1, 8
A	Dialog Information Services, File 351 WPIL, Dialog accession no. 2971640, WPI Acc no: 82-19622E/10, (HAEMATOLOGY BLOOD): "Thrombocyte concentrate prodn. for use in haemastais includes treating blood or platelet rich plasma with tetracycline hydrochloride to reduce clumping", & SU 833246, A, 810531, 8210 (Basic)	4-7
A	Dialog Information Services, File 351 WPIL, Dialog accession no. 2926038, WPI Acc No: 82-00055E/01, (SPANGENBERG P.): "Trombocyte suspension or concentrate prodn. using azo-dicarboxylic bis-dimethylamid, i.e. diamide, as re-suspension agent", & DD 151108, A, 811008, 8201 (Basic)	4-7